

Effect of Short-Term Cigarette Smoke Exposure on Body Weight, Appetite and Brain Neuropeptide Y in Mice

Hui Chen¹, Ross Vlahos^{2,3}, Steve Bozinovski^{1,3}, Jessica Jones^{1,3}, Gary P Anderson^{1,2,3} and Margaret J Morris^{*,1}

¹Department of Pharmacology, The University of Melbourne, Victoria, Australia; ²Department of Medicine, The University of Melbourne, Royal Melbourne Hospital, Victoria, Australia; ³CRC for Chronic Inflammatory Diseases, The University of Melbourne, Victoria, Australia

Although nicotinic receptors have been demonstrated in hypothalamic appetite-regulating areas and nicotine administration alters food intake and body weight in both animals and humans, the mechanisms underlying the effects of smoking on appetite circuits remain unclear. Conflicting effects of nicotine on the major orexigenic peptide, neuropeptide Y (NPY), have been observed in the brain, but the effects of smoking are unknown. Thus, we aimed to investigate how cigarette smoking affects body weight, food intake, plasma leptin concentration, hypothalamic NPY peptide, adipose mass and mRNA expression of uncoupling proteins (UCP), and tumor necrosis factor (TNF) α . Balb/C mice (8 weeks) were exposed to cigarette smoke (three cigarettes, three times a day for 4 consecutive days) or sham exposed. Body weight and food intake were recorded. Plasma leptin and brain NPY were measured by radioimmunoassay. UCPs and TNF α mRNA were measured by real-time PCR. Food intake dropped significantly from the first day of smoking, and weight loss became evident within 2 days. Brown fat and retroperitoneal white fat masses were significantly reduced, and plasma leptin concentration was decreased by 34%, in line with the decreased fat mass. NPY concentrations in hypothalamic subregions were similar between two groups. UCP1 mRNA was decreased in white fat and UCP3 mRNA increased in brown fat in smoking group. Short-term cigarette smoke exposure led to reduced body weight, food intake, and fat mass. The reduction in plasma leptin concentration may have been too modest to increase NPY production; alternatively, change in NPY or its function might have been offset by nicotine or other elements in cigarette smoke.

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INTRODUCTION

An inverse relationship between cigarette smoking and body weight has been documented in many studies. Smokers weigh less than nonsmokers of the same age and gender, and anorexia often occurs with cigarette smoking (Albanes *et al*, 1987; Klesges *et al*, 1989; Perkins, 1992). Cessation of smoking without nicotine replacement therapy is usually accompanied by weight gain (Stamford *et al*, 1986; Levin *et al*, 1987; Williamson *et al*, 1991; O'Hara *et al*, 1998). As a consequence, some people are willing to use smoking in order to lose or maintain their body weight, and are reluctant to stop smoking due to concerns about weight

gain on cessation, especially among younger smokers (Crisp *et al*, 1999; Wee *et al*, 2001). The effects of cigarette smoking on appetite may also contribute to the morbidity and mortality of smokers with chronic lung disease and wasting.

This action of smoking on body weight appears to be nicotine mediated as indicated by Hajek *et al* (1988). Previous studies in both humans and animals have reported that nicotine administration decreases body weight and caloric intake (Wager-srdar *et al*, 1984; Grunberg *et al*, 1986; Hajek *et al*, 1988; Bellinger *et al*, 2003; Bishop *et al*, 2004). Thus, the effects of nicotine to suppress appetite and decrease food intake leading to reduced body weight are not confined to human subjects.

The brain responds to altered energy homeostasis by adjusting food intake. Neuropeptides in the hypothalamus play a pivotal role in regulating food intake, and their actions in this region are known to be modulated by circulating factors such as leptin. Neuropeptide Y (NPY), a 36-amino-acid neuropeptide, which is abundant in the mammalian brain and highly concentrated in the hypothalamus, is known to play a central role in hypothalamic mechanisms, which increase food intake (Woods *et al*, 1998). Nicotine binding sites have been demonstrated in

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*Correspondence: Assoc Professor Dr MJ Morris, Department of Pharmacology, The University of Melbourne, Victoria 3010, Australia, Tel: 61 3 8344 5745, Fax: 61 3 8344 0241,

E-mail: mjmorris@unimelb.edu.au

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appetite-regulating regions of the hypothalamus, suggesting that centrally mediated actions of nicotine may contribute to the reduced appetite and body weight loss (Jo *et al*, 2002). If NPY signaling is a possible target for nicotine's anorexic effects, a suppression of NPY expression might be expected with nicotine treatment. Frankish and co-workers found that both acute (24 h) and longer-term (2 week) nicotine injection decreased NPY content and NPY mRNA expression in the arcuate nucleus (Arc) and paraventricular nucleus (PVN) of the rat; however, Li and co-workers have shown a positive correlation between NPY mRNA expression and the dose of nicotine used (Frankish *et al*, 1995; Li *et al*, 2000). Thus, the picture with regard to the effects of nicotine on NPY in the central nervous system is unclear. Further, the effects of smoking, rather than nicotine, on brain mediators involved in appetite regulation, have not been investigated to date.

In addition to examining NPY, we also investigated the effects of smoking on other markers of obesity and energy homeostasis, such as leptin and uncoupling proteins (UCP). Leptin is a hormone secreted by adipose tissue that is known to act centrally to inhibit the effects of NPY, apparently by inhibiting its synthesis in the Arc (Stephens *et al*, 1995; Cusin *et al*, 1996; Erickson *et al*, 1996; Elmquist *et al*, 1999). The distribution of leptin receptors was found to overlap with that of nicotinic receptors in the hypothalamus, indicating a possible link between these two factors (Jo *et al*, 2002). UCPs are mitochondrial carrier proteins, which are able to dissipate the proton gradient of the inner mitochondrial membrane. This uncoupling process reduces the amount of ATP generated through oxidation of fuels and increases thermogenesis, which is related to energy metabolism (Dalgard and Pedersen, 2001).

Smoking exerts an inflammatory stimulus on macrophages, which brings about the production of inflammatory cytokines, such as tumor necrosis factor (TNF) α , which might be an important early event in the development of disease states associated with smoking (Fernandez-Real *et al*, 2003). Increased TNF α activity (Fernandez-Real *et al*, 2003) and decreased actions have been observed in smokers and nicotine administration studies (Ouyang *et al*, 2000; Liu *et al*, 2001). Produced by both immunocompetent cells and adipocytes, TNF α can regulate lipid metabolism, adipocyte differentiation, inhibit appetite, and is a mediator of cachexia (Torti *et al*, 1989; Spiegelman and Hotamisligil, 1993; Ventre *et al*, 1997; Bullo-Bonet *et al*, 1999; Langhans and Hrupka, 1999). Therefore, we also determined the effect of cigarette smoke exposure on TNF α expression in adipose tissue.

Therefore, the aim of this study was to measure the changes in food intake, body weight, hypothalamic NPY content, and expression of UCP1, UCP3, and TNF α mRNA in fat tissue in response to short-term (4 days) cigarette smoke exposure in mice.

MATERIALS AND METHODS

Animals

Development of respiratory disease following cigarette smoke exposure is strain dependent (Guerassimov *et al*, 2004), and in this study, Balb/C mice were selected based on

the inflammatory and cytokine responses in related respiratory studies in our laboratory. Male Balb/C mice (aged 7 weeks, $n=32$) were obtained from the Animal Resource Centre Pty Ltd (Perth, Australia), and were housed at $20\pm2^{\circ}\text{C}$ in microisolator cages, and maintained on a 12:12 h light/dark cycle (lights on at 0600). They were allowed *ad libitum* access to standard rodent chow and water. Mice were allowed a week to adapt to their new environment. Animals were monitored daily; food intake per cage and individual body weight were measured three times per week during this period. After acclimatization, mice were randomly divided into two groups, cigarette smoke exposed (smoking group) or sham exposed (control group). The animals exposed to cigarette smoke were placed inside a perspex chamber (18 l) and exposed to the smoke produced by three cigarettes (Winfield Red, 16 mg or less of tar, 1.2 mg or less of nicotine, and 15 mg or less of CO), three times (0900, 1200 and 1500) a day for 4 consecutive days. Control mice were handled similarly without smoke exposure. Food intake and body weight were measured daily. All procedures were approved by the Animal Experimentation Ethics Committee of The University of Melbourne.

Sample Collection

On the 5th day, mice were anesthetized (ketamine/xylazine 15/30 mg/kg, intraperitoneal), and blood was collected from the abdominal vena cava into heparinized tubes (5000 IU/ml; Fisons, Australia), and centrifuged at 10 000 r.p.m. and 4°C for 8 min. Separated plasma was stored at -80°C for subsequent determination of plasma leptin and corticosterone concentrations. Mice were decapitated and the brain removed and rapidly dissected on ice into regions containing PVN, Arc, anterior and posterior hypothalamus (AH and PH), and medulla. Brain regions were weighed and stored at -80°C for later determination of NPY peptide content. Body fat (brown adipose tissue (BAT), left retroperitoneal white adipose tissue (RpWAT), testicular WAT) and liver were dissected and weighed. BAT and RpWAT were stored at -80°C for later measurement of mRNA of UCP1, UCP3, and TNF α .

Brain NPY, Plasma Leptin, and Corticosterone Assays

Endogenous NPY from the various brain regions was extracted by boiling the tissues in 0.5 M acetic acid, homogenization by hand with a glass homogenizer, and centrifugation at 7500 r.p.m. for 30 min at 4°C . The supernatant was decanted, and 50 μl samples were lyophilized and reconstituted with assay buffer (0.04 M sodium phosphate buffer containing 0.01 M EDTA, 0.1 M NaCl, 0.02% NaN_3 , 0.25% BSA (pH 7.3)). NPY-like immunoreactivity in the various brain regions was measured by a specific radioimmunoassay developed in our laboratory using synthetic NPY as standard (10–1280 pg/tube, Auspep, Australia) (Morris *et al*, 1986). Samples were incubated with NPY antibody overnight at 4°C . [^{125}I]NPY labeled with Bolton and Hunter reagent (3000 Ci/mmol, Amersham, Australia) was added and the incubation continued overnight. Bound and free radioligand were separated by the addition of nonimmunized rabbit serum and sheep anti-

rabbit second antibody followed by centrifugation at 3000 r.p.m. at 4°C for 35 min (RT7, Sorvall instruments). The bound fraction was then counted in a gamma counter. The detection limit for the radioimmunoassay was routinely 2 pg NPY/tube and the intra- and interassay coefficients of variation were 6 and 13%, respectively. NPY in each brain region was calculated as ng NPY/mg tissue. Plasma leptin and corticosterone concentrations were measured using a commercially available radioimmunoassay kit (Linco, MI, USA, and MP Biomedicals Europe, Belgium, respectively).

UCP1, UCP3, and TNF α Measurement

Total RNA was isolated from 10 mg of both WAT and BAT using an RNAeasy kit (Qiagen) according to the manufacturer's instructions. The purified total RNA rep was used as a template to generate first-strand cDNA synthesis using Super Script II (Invitrogen) as described previously (Bozinovski *et al*, 2004). The reaction mix containing 1 μ g of RNA, 250 ng of random hexamers (Promega), and 10 mM dNTP mix was diluted to 12 μ l in sterile water, heated to 65°C for 5 min and chilled on ice for 1 min. First-strand synthesis was then performed in a 20 μ l total reaction volume by adding 50 mM Tris-HCl (pH 8.3), 75 mM KCl, 3 mM MgCl₂, 10 mM DTT, 40 U RNaseout and 200 U Superscript II reverse transcriptase enzyme at 42°C for 50 min. The reaction was inactivated by heating at 70°C for 15 min. cDNA was diluted 10-fold in sterile water and stored at -20°C prior to amplification. Quantitative real-time PCR was performed as described previously (ABI PRISM 7900 HT Sequence Detection System) (Bozinovski *et al*, 2004) using predeveloped primers from Applied Biosystem. Briefly, gene expression was quantified by multiplexing single reaction, where our gene of interest (UCP1, UCP3, and TNF α) was standardized to control (18s rRNA). An individual BAT sample from the control group was then arbitrarily assigned as a calibrator against which all other samples are expressed as a fold difference.

Statistical Analyses

Results are expressed as mean \pm SEM. Body weight of smoke-exposed and control mice were analyzed using analysis of variance with repeated measures, followed by a *post hoc* least significance difference test. Difference in daily food intake, fat and organ mass, plasma leptin and corticosterone concentrations, brain NPY concentration and content, and relative expression of mRNA were analyzed using Student's unpaired *t*-tests. The relation between plasma leptin concentration and body weight, body weight gain and WAT was examined using simple linear regression.

RESULTS

Both food intake and body weight of the two experimental mice groups were well matched at the beginning of this study as shown by Figure 1a and b, respectively. Over the whole experimental period, daily food intake of control mice was stable. After the first day of cigarette smoke exposure, food intake was somewhat lower than control mice, and this difference increased over time. From the

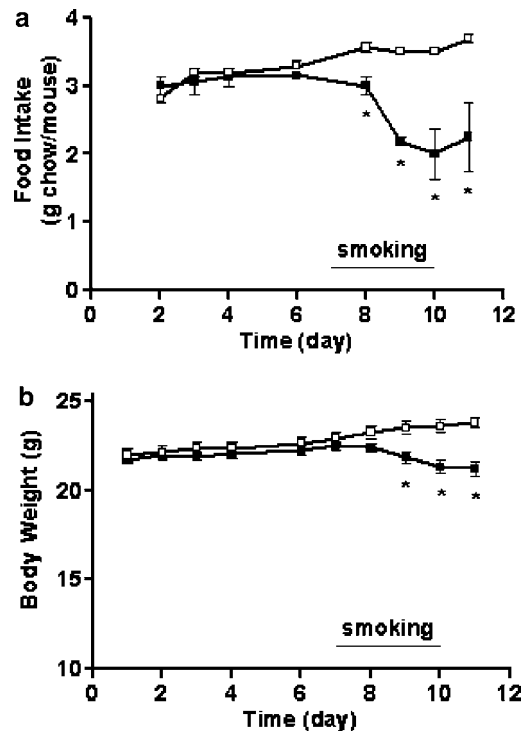


Figure 1 (a) Food intake (g chow/mouse/day) of control (open bar) and smoking (filled bar) Balb/C mice during experimental period. Mice were exposed to cigarette smoke or sham exposed from day 7 to 10. Results are expressed as mean \pm SEM of 16 mice in each group. *Significant difference between treatment groups ($p < 0.05$). (b) Body weight of control mice (open bar, $n = 16$) and smoking mice (filled bar, $n = 16$) during the experimental period. Mice were exposed to cigarette smoke or sham exposed from day 7 to 10. Results are expressed as mean \pm SEM. *Significant difference between treatment groups ($p < 0.05$).

second day of smoke exposure, food intake was reduced to 60% of the control level (Figure 1a, $p < 0.05$). The total 4 day chow intake of the mice exposed to cigarette smoke was reduced by 34% (9.4 vs 14.3 g/mouse in control mice, $p < 0.05$). There was a time effect on the growth of control mice, reflecting weight gain, which was opposite in the smoking group ($p < 0.05$, Figure 1b). Body weight of the smoking group was significantly reduced within 2 days of cigarette smoking (21.9 ± 0.3 vs 23.6 ± 0.3 g for smoking and control group, respectively, $p < 0.05$, Figure 1b) and pretreatment (day 7) levels ($p < 0.05$, Figure 1b). After 4 days of cigarette smoke exposure, the average body weight of the smoking group was 10% less than the control group ($p < 0.05$, Table 1).

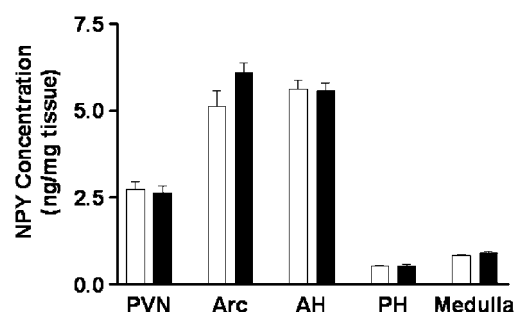
Animals exposed to 4 days of cigarette smoke displayed lower weights of the liver, BAT, and RpWAT relative to control mice ($p < 0.05$, Table 1). Notably for BAT and RpWAT, these differences were still pronounced when data were calculated as a percentage of body weight (BAT 0.38 ± 0.02 vs $0.47 \pm 0.02\%$; RpWAT 0.48 ± 0.04 vs $0.59 \pm 0.03\%$, in smoking and control group, respectively, $p < 0.05$). Plasma leptin concentrations in the smoking group decreased significantly, in line with the decreased white fat mass ($p < 0.05$, Table 1). When both groups were combined, plasma leptin concentration was significantly correlated with WAT, body weight, and body weight gain ($r = 0.60, 0.54$, and 0.58 , respectively, $p < 0.05$, $n = 15$). No

Table 1 Effect of Cigarette Smoke Exposure on Body Weight, Liver, Adipose Tissue Mass, Plasma Leptin, and Corticosterone Concentration

	Control	Smoke exposure
Body weight (pre-exposure) (g)	23.0 ± 0.3	22.5 ± 0.2
Body weight (postexposure) (g)	23.6 ± 0.3	21.3 ± 0.4*
Liver (mg)	1188.5 ± 38.5	929.8 ± 20.3*
BAT (mg)	110.7 ± 6.5	80.0 ± 4.0*
RpWAT (mg)	139.1 ± 7.7	104.3 ± 9.7*
Testicular WAT (mg)	471.7 ± 24.9	416.4 ± 34.9*
Leptin (ng/ml)	6.5 ± 0.5	4.3 ± 0.5*
Corticosterone (ng/ml)	212.4 ± 50.7	247.3 ± 33.6

Results are expressed as mean ± SEM (for body weights, $n = 32$; for tissue masses, $n = 11$, 12 , respectively; for plasma hormone concentrations, $n = 8$, 7 , respectively).

*Significantly different from control mice ($P < 0.05$).

**Figure 2** NPY concentration of brain regions of control mice (open bar, $n = 8$) and smoking mice (filled bar, $n = 7$) after 4 days cigarette smoke exposure. Results are expressed as mean ± SEM.

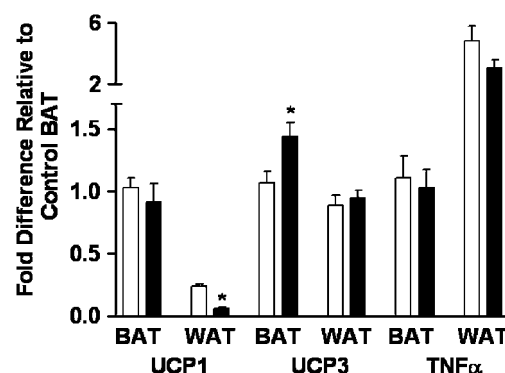
difference in plasma corticosterone levels was observed between treatment groups at the end of the experiment (Table 1).

NPY concentrations of the hypothalamic regions, AH, PH, PVN, and Arc, as well as medulla, were not different between the control and the smoking groups at the end of the experiment (Figure 2). The total hypothalamic NPY content was not altered by 4 days smoke exposure (smoking group 82.3 ± 1.9 ng vs control group 86.2 ± 2.9 ng).

UCP1 mRNA expression was decreased by 76% in WAT in the smoking group, but no difference was found in BAT between the smoking and control group. BAT UCP3 mRNA expression was significantly increased in cigarette smoke-exposed mice, while no change was observed in WAT. TNF α mRNA expression was not affected by cigarette smoking in either WAT or BAT (Figure 3).

DISCUSSION

In this study, we examined the hormonal, neurochemical, and metabolic changes induced by short-term (4 consecutive days) cigarette smoke exposure in mice. Using this protocol, we previously determined blood carboxyhemoglobin content to be 3% (unpublished observation),

**Figure 3** UCP1, UCP3, and TNF α mRNA expression in BAT and WAT of control mice (open bar, $n = 7$) and smoking mice (filled bar, $n = 8$) after 4 days cigarette smoke exposure, standardized to 18s rRNA, expressed relative to a BAT sample from the control group (fold difference). Results are expressed as mean ± SEM. *Significantly different from control group ($p < 0.05$).

suggesting a moderate level of smoke exposure in the smoking group, compared to 10–14% in heavy smokers (Benowitz *et al*, 1982). When exposed to cigarette smoke, the mice showed a consistent reduction in chow intake compared to both their baseline levels before cigarette smoke exposure was implemented, and the chow intake of the control group. The unchanged levels of plasma corticosterone, a stress response hormone, between experimental groups, suggest that decreased food intake might not be due to any greater stress related to smoke exposure. This reduced appetite with smoke exposure is consistent with the anorexia commonly observed among human smokers and previous studies using nicotine administration (Grunberg *et al*, 1986; Albanes *et al*, 1987; Levin *et al*, 1987; Hajek *et al*, 1988; Klesges *et al*, 1989; Bellinger *et al*, 2003; Bishop *et al*, 2004). However, clearly, it is difficult to draw comparisons between the experiments using nicotine administration and effects of cigarette smoke, which contains multiple factors, and can cause inflammatory changes, which might alter energy metabolism. The decreased food intake most likely led to the significant reduction in body weight, which was 10% less than control mice at death, reflected by a significant reduction in adipose tissue, particularly white fat masses. When data were calculated as a percentage of body weight, white fat masses remained significantly reduced in the smoking group, indicating that decreased body fat largely contributed to the reduced body weight, and suggesting that fat deposits might be used as an energy supply under these conditions of negative energy balance. The anorexia and weight loss caused by cigarette smoke exposure in mice appears to resemble the effects of cigarette smoking in humans.

NPY is synthesized in the Arc, from where cells project to the PVN and other hypothalamic areas (Bai *et al*, 1985; Chronwall *et al*, 1985). Exogenous administration of NPY into the cerebroventricle results in a dramatically increased carbohydrate and fat intake and decreased energy expenditure in rodents, and eventually, to a state of obesity (Zarjevski *et al*, 1993). NPY peptide production is increased with starvation, food restriction and in genetically obese

animals (eg *ob/ob* mice), and decreased with refeeding, suggesting that NPY may be a critical mediator for the maintenance of body weight (Beck *et al*, 1990; Brady *et al*, 1990; Wilding *et al*, 1993; Swart *et al*, 2002). Since nicotinic receptors have been demonstrated in the appetite-regulating area of the hypothalamus, cigarette smoking might change appetite in mice by affecting the energy homeostasis circuits including both those releasing orexigenic peptides, such as NPY and melanin concentrating hormone, and anorexic peptides, such as cocaine- and amphetamine-regulated transcript and pro-opiomelanocortin (Jo *et al*, 2002). However, NPY concentrations in the hypothalamic subregions, AH, PVN, PH, and Arc, as well as the medulla, were not affected by 4 days of smoke exposure. In previous nicotine administration studies in rat, decreased NPY peptide in PVN and Arc was observed after 24 h of high-dose (12 mg/kg/day) nicotine administration that reduced food intake by 30% (Frankish *et al*, 1995), while food intake of mice exposed to cigarette smoke in the present study was only 16% less than control mice in the first 24 h, suggesting the extent of negative energy status induced by the cigarette smoke exposure we used might be insufficient to regulate hypothalamic NPY. However, in another study, hypothalamic NPY peptide was found to be increased after 14 days of low-dose (4 mg/kg/day) nicotine treatment, which reduced food intake by 19.5% in rats (Li *et al*, 2000), suggesting that the present treatment may have been too short to cause significant changes in NPY peptide. Furthermore, previously it was found that nicotine can directly inhibit the hyperphagia produced by exogenous administration of NPY into the PVN (Bishop *et al*, 2002), suggesting it is possible that the effects of brain NPY can be offset by nicotine or other elements in cigarette smoke. The effects of nicotine administration on NPY peptide in mice have not been documented previously. Decreased leptin levels would normally decrease the inhibitory effects of leptin on NPY and thus increase NPY production (Stephens *et al*, 1995; Elmquist *et al*, 1999). Although we have previously described significant negative correlations between plasma leptin and hypothalamic NPY in a number of paradigms including weight loss following vagotomy (Furness *et al*, 2001), and chronic high fat diet (Hansen *et al*, 2004), the impact of cigarette smoke exposure on leptin here was relatively modest compared to these interventions.

In addition to the decreased food intake, it is possible that other factors involved in energy balance may also have contributed to the decreased body weight observed in mice exposed to cigarette smoke. While it is generally accepted that UCP1 is exclusively expressed in BAT in rodents, it can be induced in WAT at a lower levels than in BAT (Nagase *et al*, 1996; Yoshida *et al*, 1999). In our study, UCP1 in BAT was not affected by 4 days smoke exposure, suggesting BAT thermogenesis was largely unaffected by cigarette smoking in this protocol. By using quantitative real-time PCR, UCP1 mRNA can be detected under basal conditions in WAT in mice. The expression of UCP1 mRNA in WAT was found to be significantly decreased by cigarette smoke exposure. However previously, it was reported that UCP1 mRNA can be induced in both BAT and WAT by nicotine treatment, probably enhancing the energy expenditure in this situation (Yoshida *et al*, 1999; Arai *et al*, 2001). These changes

highlight the need to examine metabolic effects of smoking *per se* rather than nicotine administration. Fasting and chronic food deprivation can downregulate UCP1 expression in BAT (Samec *et al*, 1998; Sivitz *et al*, 1999), with changes in WAT unknown. However, in our study, a 34% reduction of total food intake over 4 days cigarette smoke exposure did not affect UCP1 mRNA expression in BAT, but reduced it in WAT. Although overexpression of UCP1 mRNA in WAT can cause obesity resistance (Kopecky *et al*, 1996), it is not clear as to how the decreased UCP1 mRNA might contribute to the maintenance of energy balance in this situation. It is difficult to separate any effects of nicotine or food restriction on UCP1, as both have been shown to regulate expression of UCPs.

UCP3, a homolog of UCP1, is expressed in BAT and skeletal muscle and is implicated in the regulation of mitochondrial fatty acid transport and influences basal metabolic rate (Samec *et al*, 1998). In our study, UCP3 mRNA was significantly increased in BAT in the face of the reduced food intake following smoke exposure, which was opposite to previous studies where the UCP3 mRNA expression in BAT was downregulated by fasting and food deprivation, a physiological response to conserve energy expenditure (Samec *et al*, 1998; Sivitz *et al*, 1999). The role of UCP3 in lipid regulation in skeletal muscle can be extended to BAT (Samec *et al*, 1998). An increase in UCP3 would increase uncoupling of mitochondrial respiration and increase energy expenditure or heat dissipation, which suggests that in the smoke-exposed mice, lipid utilization and energy expenditure were upregulated, rather than decreasing energy expenditure to maintain homeostasis. Thus, cigarette smoke exposure probably disturbs energy homeostasis, which may crucially contribute to the weight and adipose loss observed. Very little data exist on the effects of nicotine administration on BAT UCP3 mRNA expression in rodents to date.

TNF α is increasingly recognized to be involved in the pathogenesis of obesity, and its activation seems to be associated with increased energy expenditure and weight loss (Toomey *et al*, 1995; Tracey and Cerami, 1992). Previously, it was found that TNF α is increased in patients with anorexia (Vaisman and Hahn, 1991), and exogenous administration of TNF α had inhibitory effects on food intake in rats (Fantino and Wieteska, 1993; Sonti *et al*, 1996; McCarthy, 2000). In addition, TNF α can affect lipid metabolism *in vivo* by increasing lipolysis in adipose tissue, and inhibit adipose cell differentiation *in vitro* (Torti *et al*, 1989; De clerq *et al*, 1996; Kern, 1997). However, our observation of no change in TNF α mRNA expression in both BAT and WAT suggests that TNF α (at least in adipose tissue) did not contribute to anorexia and the loss of fat deposits following cigarette smoke exposure.

In summary, 4 days cigarette smoking caused loss of appetite and body weight in mice, and accordingly negative energy balance, but increased lipid utilization and energy expenditure reflected by an induced UCP3 mRNA expression in BAT. Hypothalamic NPY concentration and content were not changed by 4 days cigarette smoke exposure, probably due to the short-term nature of this experiment or the inhibitory effects of cigarette smoke exposure. Further work is underway to determine the effects of more chronic periods of cigarette smoking.

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